Analysis of the heterogeneity of immunoglobulins in human serum Kenneth C. Parker and Stephen J. Hattan SimulTOF Systems, Sudbury, MA Poster

Background: Immunoglobulins are typically very heterogeneous in human sera. In certain disease states, the diversity becomes skewed such that certain clonal antibodies become dominant, or so that antibodies produced from certain immunoglobulin variable region families become enriched or depleted. It would be useful to get a snapshot of immunoglobulin diversity by using a protocol that is as simple as possible. This might be feasible by linear MALDI, but to identify heavy chain classes and variable region families, tryptic peptides provide relevant information. In MALDI of complex mixtures, argininecontaining (R) peptides are most readily detected, and there are relatively few R peptides from Ig constant regions. To test these ideas, we have developed methods to perform peptide mass fingerprinting and MS-MS database searching against immunoglobulin-specific protein databases (and ordinary human databases). The samples consist of unseparated protein digests, which yields information about the relative concentration of immunoglobulin variable reagion families.

Experimental System:

•Buy Human Serum from Sigma, and Goat anti hemoglobin serum.

•Purify Immunoglobulins using ProteinA membrane (see Poster

•Reduce and digest with bovine trypsin (Worthington).

• Dilute into HCCA matrix, and spot.

•Collect reflectron spectra, and MS-MS Spectra.

•Look for pattern of patterns corresponding to Ig Constant and Variable regions.

•Compare scores of best Ig sequence to the rest of human proteome. •Search against human SwissProt or human IMGT database.

• Proprietary software used for PMF and MSMS.

•For MSMS analyses, individual sequences are pooled, and for Ig, many distinct proteins likely can be digested to generate each peptide.



Fig. 1A. Tryptic digest of whole serum (from Sigma), dominated by albumin.

Cartoons of PMF identifications based on real mass spectra; mostly confirmed by multiple MS-MS spectra from the same (or similarly prepared digests). Note that many other peptides can be mapped at mz values higher than shown in the figure. Most strong signals correspond to Ig constant regions (including IgA constant regions), but many signals are clearly derived from variable region peptides. Some strong signals have not yet been explained by expected peptides from serum (or trypsin or keratin).

by Hattan).

Fig. 2 Inset of parent spectrum between mz 1864 and 1924.



This region contains most theoretical N-terminal R-containing peptides from Ig H, IgK and IgL. The above histogram counts the number of distinct variable R-containing peptides that match within 20 ppm of the measured mass from a human germ line protein sequence library. (Many match < 5 ppm). Favorable constant region peptides were granted a histogram score of 5 to emphasize that they are expected to be present. Some measured masses apparently correspond to unique germ line encoded peptides. Other masses may well correspond to multiple variable region peptides with slightly different masses. In all case, it is expected that there are many other precursor peptides from additional N-temrinal variable region peptides with mutations, and also from variable region and constant peptides from other regions. The weak signal at 1902 is puzzling; additional work is needed to determine whether an expected constant region peptide is low in abundance, or perhaps incomplettely digested.

PMF proposed IDs from whole serum digest. Table 1.

N	Symb	Length	Name	Matches	Score	%IM	ppw
1	ALB	609	Serum albumin	18	408634	41.2	5.8
2	IGHA1	353	Ig alpha-1 chain C	4	208877	4.7	2.3
- 3	APOA1	267	Apolipoprotein A-I	11	35225	6.0	8.0
4	A2M	1474	Alpha-2-macroglobulin	11	30480	3.6	5.9
5	IGHM	452	Ig mu chain C	5	28292	1.4	3.8
6	wrong	140	Putative	4	16000	0.9	4.6
7	C3	1663	Complement C3	15	14097	3.2	6.7
8	wrong	70	NADH dehydrogenase	3	10163	3.6	4.5
9	wrong	204	Rho GDP-dissociation	2	6773	2.3	5.2
10	TRFE_hu	698	serotransferrin	7	4705	2.9	8.9

Proteins in green are abundant serum proteins. The others are wrong, and this demonstrates the limit of mass fingerprinting from complex mixtures like whole serum. Slightly different sample preparations typically yield similar numbers of well known serum proteins, but the exact proteins in these lists is variable. Of course, if albumin is not the top hit, something is very wrong! Immunoglobulins have a disadvantage in PMF because prominent peptides are listed separately in the database.

Fig. 3 MSMS spectrum of parent mass 1895.989



MSMS spectrum

S	Search human Ig database from IMGT with 25 amuy window -> 1865 peptides; top 40 are IgV N-termini									
I		р	MassTheo	MassExp	ppm	Sequence	M	TrS	Symb	Name
	1	. 0	1897.00	1895.99	-535.0	EVQLLESGGGLVEPGGSLR	28	1200	IGHV	IGHV3-23*01
	2	0	1896.06	1895.99	-35.2	EVQLLESGGGLVKPGGSLR	28	1179	IGHV	IGHV3-21*01
	3	1	1896.06	1895.99	-35.2	EVKLLESGGGLVQPGGSLR	28	1157	IGHV	IGHV3-66*01
	4	0	1896.02	1895.99	-16.0	EVQLLESGGGLVQPGGSLR	28	1157	IGHV	IGHV3-23*01
	5	0	1895.04	1895.99	503.0	EVQLLQSGGGLVQPGGSLR	28	1093	IGHV	IGHV3-23*01
	6	0	1896.06	1895.99	-35.2	EVQLVETGGGLVKPGGSLR	30	1015	IGHV	IGHV3-30*02
	7	0	1896.02	1895.99	-16.0	EVQLVETGGGLVQPGGSLR	30	1015	IGHV	IGHV3-53*02
	8	0	1896.98	1895.99	-521.7	EVQQVESGGGLVQPGGSLR	28	1007	IGHV	IGHV3-23*04
	9	0	1887.92	1895.99	4253.9	EVQLSESGGGMVQPGGSLR	29	981	IGHV	IGHV3-23*01
	10	0	1897.96	1895.99	-1040.7	EVQLVESGGGLDQPGGSLR	26	968	IGHV	IGHV3-7*01
	11	0	1885.94	1895.99	5297.8	EVQMVDSGGGLVQPGGSLR	24	928	IGHV	IGHV3-7*01
	12	0	1896.06	1895.99	-35.1	EVQLVESGGALVKPGGSLR	29	897	IGHV	IGHV3-15*02
	13	0	1896.02	1895.99	-16.0	EVQLVESGGALVQPGGSLR	29	880	IGHV	IGHV3-7*01
	14	0	1896.02	1895.99	-16.0	EVQLVESGGAVIQPGGSLR	29	871	IGHV	IGHV3-43*01
	15	0	1896.02	1895.99	-16.0	EVQLVESAGGLVQPGGSLR	28	850	IGHV	IGHV3-15*08
	16	0	1896.06	1895.99	-35.2	EVQLVESGGGLLKPGGSLR	28	787	IGHV	IGHV3-15*01
	17	0	1896.06	1895.99	-35.2	EVQLVESGGGLIKPGGSLR	28	787	IGHV	IGHV3-21*01
	18	0	1896.02	1895.99	-16.0	EVQLVESGGGLLQPGGSLR	28	756	IGHV	IGHV3-74*01
	19	0	1896.02	1895.99	-16.0	EVQLVESGGGLIQPGGSLR	28	756	IGHV	IGHV3-53*01
	20	0	1887.95	1895.99	4240.7	DIVMTQSPGTLSLSPGER	24	750	IGKV	IGKV3-20*01
	21	0	1896.02	1895.99	-16.0	EVQLVESGGVLAQPGGSLR	27	729	IGHV	IGHV3-74*01
	22	0	1896.00	1895.99	-6.6	qVQLMESGGGLVKPGGSLR	23	727	IGHV	IGHV3-11*01
	23	0	1896.00	1895.99	-6.6	eVQLMESGGGLVKPGGSLR	23	727	IGHV	IGHV3-15*07
	24	0	1905.02	1895.99	-4763.0	EVHLLESGGGLVQPGGSLR	20	703	IGHV	IGHV3-23*01
	25	1	1896.06	1895.99	-35.2	VKLLEESGGGLVQPGGSLR	23	695	IGHV	IGHV3-23*01
	26	0	1895.07	1895.99	483.8	QVQLLESGGGLVKPGGSLR	23	694	IGHV	IGHV3-11*03
	27	1	1898.00	1895.99	-1059.9	EVKVEESGGGLVQPGGSLR	25	688	IGHV	IGHV3-66*01
	28	0	1896.03	1895.99	-21.9	EVQLVESGGGVVRPGGSLR	26	687	IGHV	IGHV3-20*01
	29	0	1895.99	1895.99	-2.7	QVQLVQSGGGVEQPGGSLR	25	687	IGHV	IGHV3-48*03
	30	1	1897.05	1895.99	-560.1	QITLKESGGGLVQPGGSLR	23	679	IGHV	IGHV3-48*03
	31	1	1896.09	1895.99	-54.4	VKLLEESGGGLVKPGGSLR	23	679	IGHV	IGHV3-23*01
	32	0	1895.97	1895.99	12.6	eVQLMESGGGLVQPGGSLR	23	679	IGHV	IGHV3-15*07
	33	0	1895.04	1895.99	503.0	QVQLLESGGGLVQPGGSLR	23	649	IGHV	IGHV3-23*01
	34	1	1895.07	1895.99	483.8	VKLLEQSGGGLVQPGGSLR	23	646	IGHV	IGHV3-23*01
	35	1	1895.07	1895.99	483.8	QVKLLESGGGLVQPGGSLR	23	633	IGHV	IGHV3-23*01
	36	1	1895.11	1895.99	464.6	QVKLLESGGGLVKPGGSLR	23	618	IGHV	IGHV3-21*01
	37	0	1896.00	1895.99	-6.6	qMQLVESGGGLVKPGGSLR	21	606	IGHV	IGHV3-11*03
	38	0	1896.00	1895.99	-6.6	eMQLVESGGGLVKPGGSLR	21	606	IGHV	IGHV3-15*01
	39	0	1895.07	1895.99	483.8	VQLLEQSGGGLVKPGGSLR	22	589	IGHV	IGHV3-15*01
	40	0	1896.02	1895.99	-16.0	VQLLEESGGGLVQPGGSLR	22	589	IGHV	IGHV3-7*01
2	Sear	ch h	uman data	base with	25 amu wir	ndow -> 19999 peptides; best h	it score 656	-		
I		р	MassTheo	MassExp	ppmP	Sequence	M	TrS	Symb	Name
	1	0	1884.96	1895.99	5815.5	GVCISPEAIVTDLPEDVK	25	656	wtt13	Uncharacterized
	1	7 X				······································	г г Л		I A 117 19	

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I		р	MassTheo	MassExp	рртР	Sequence
	1	0	1884.96	1895.99	5815.5	GVCISPEAIVTDLPEDV
	2	0	1889.96	1895.99	3180.5	PRPPPPPPPPEESCPA
	3	1	1887.96	1895.99	4232.7	SDCPLGVTKEIAIWAER
	4	2	1903.04	1895.99	-3716.8	TPSTASLTRTPSRASLTF
	5	1	1902.89	1895.99	-3638.1	QDIMPEVDKQSGSPES

32 distinct variable region peptides scored higher than any peptides from the human proteome, based on 40,000 human proteins in SwissProt, which does not contain all common Ig variable sequences from the germ line. Many of the strongest MSMS peaks can be explained by ions shared by many of these peptides. Note that each peptide above may be common to many germ line genes; other are specific to individual V region families, barring back mutation.

Conclusions:

- •Can see peptides from Ig in tryptic digest of whole serum.
- •Can confirm some of them by MSMS without separation (not shown).
- •Can readily see variable peptides in digests of Protein A purified protein.
- •Can deduce something about IgG class usage from unseparated digests.
- •Can deduce something about variable region family usage.
- •Should be useful for characterizing monoclonal gammopathies (not shown
- here).
- •In the future, study reproducibility of patterns from individuals.



